



PrimeSurface® Publication List

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Publications from 2019 to Apr. 2020 (80)

PrimeSurface® Stem Cell Differentiation Research Publications (41)

< Retinal Research and Eye disease >

1. K. Eastlake, et al, Phenotypic and Functional Characterization of Müller Glia Isolated from Induced Pluripotent Stem Cell-Derived Retinal Organoids: Improvement of Retinal Ganglion Cell Function upon Transplantation. *Stem Cells Translational Medicine*, 2019, 8.8: 775–784 [MS-9096V]
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6646702/>
2. S. Kitahata, et al, Critical Functionality Effects from Storage Temperature on Human Induced Pluripotent Stem Cell-Derived Retinal Pigment Epithelium Cell Suspensions. *Scientific Reports*, 2019, 9:2891 [MS-9096U, M or V]
<https://www.nature.com/articles/s41598-018-38065-6>
3. C. B. Mellough, et al, Systematic Comparison of Retinal Organoid Differentiation from Human Pluripotent Stem Cells Reveals Stage Specific, Cell Line, and Methodological Differences. *Stem Cells Translational Medicine*, 2019, 8.7: 694–706 [MS-9096V]
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6591558/>
4. H. C. Suen, et al, Transplantation of Retinal Ganglion Cells Derived from Male Germline Stem Cell as a Potential Treatment to Glaucoma. *Stem Cells and Development*, 2019, 28.20:1365-1375 [MS-9096U]
<https://pubmed.ncbi.nlm.nih.gov/31580778/>
5. A. Kuwahara, et al, Preconditioning the Initial State of Feeder-free Human Pluripotent Stem Cells Promotes Self-formation of Three-dimensional Retinal Tissue. *Scientific Reports*, 2019, 9: 18936. [MS-9096V]
<https://www.nature.com/articles/s41598-019-55130-w>

< Neuroscience Research >

1. H. Takeuch, et al, A scaffold-free Bio 3D nerve conduit for repair of a 10-mm peripheral nerve defect in the rats. *Microsurgery*, 2020, 40.2: 207-216. [MS-9096U, M or V]
<https://onlinelibrary.wiley.com/doi/abs/10.1002/micr.30533>
2. S. Mitsuzawa, et al, The efficacy of a scaffold-free Bio 3D conduit developed from autologous dermal fibroblasts on peripheral nerve regeneration in a canine ulnar nerve injury model: a preclinical proof-of-concept study. *Cell transplantation*, 2019, 28.9-10: 1231-1241. [MS-9096U, M or V]
<https://journals.sagepub.com/doi/full/10.1177/0963689719855346>
3. D. Lukmanto, et al, Dynamic changes of mouse embryonic stem cell-derived neural stem cells under in vitro prolonged culture and hypoxic conditions. *Stem cells and development*, 2019, 28.21: 1434-1450. [MS-9096U, M or V]
<https://www.liebertpub.com/doi/abs/10.1089/scd.2019.0101>
4. T. Sunohara, et al, MicroRNA-based separation of cortico-fugal projection neuron-like cells derived from embryonic stem cells. *Frontiers in neuroscience*, 2019, 13: 1141. [MS-9096U]

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<https://www.frontiersin.org/articles/10.3389/fnins.2019.01141/full>

5. H. Sakaguchi, et al, Self-organized synchronous calcium transients in a cultured human neural network derived from cerebral organoids. *Stem Cell Reports*, 2019, 13.3: 458-473. [MS-9096V]
<https://www.sciencedirect.com/science/article/pii/S2213671119301973>
6. W. Zhu, et al, Precisely controlling endogenous protein dosage in hPSCs and derivatives to model FOXG1 syndrome. *Nature communications*, 2019, 10.1: 1-18. [MS-9096VZ]
<https://www.nature.com/articles/s41467-019-08841-7>
7. S. N. Sarma, et al, Tyroxine hydroxylase-positive neuronal cell population is increased by temporal dioxin exposure at early stage of differentiation from human embryonic stem cells. *International journal of molecular sciences*, 2019, 20.11: 2687. [MS-9096U]
https://www.mdpi.com/1422-0067/20/11/2687/htm?_cf_chl_managed_tk=&kD2hplaZCj1mntbu9quEZZbhq.fr0pkTEHaB7DK7hSc-1641534247-0-gaNycGzNB-U
8. A. Hirota, et al, The nucleosome remodeling and deacetylase complex protein CHD4 regulates neural differentiation of mouse embryonic stem cells by down-regulating p53. *Journal of Biological Chemistry*, 2019, 294.1: 195-209. [MS-9096U, M or V]
<https://www.jbc.org/content/294/1/195.full.pdf>
9. B. Samata, et al, L1CAM is a marker for enriching corticospinal motor neurons in the developing brain. *Frontiers in cellular neuroscience*, 2020, 14: 31. [MS-9096U, M or V]
<https://www.frontiersin.org/articles/10.3389/fncel.2020.00031/full>

< Cardiomyocytes and Heart Research >

1. T. Kitsuka, et al, 2-Cl-C.OXT-A stimulates contraction through the suppression of phosphodiesterase activity in human induced pluripotent stem cell derived cardiac organoids. *PLoS ONE*, 2019, 14.7: e0213114 [MS-9096V]
<https://journals.plos.org/plosone/article/file?type=printable&id=10.1371/journal.pone.0213114>
2. S. Yoshida, et al, Hydrogel Microchambers Integrated with Organic Electrodes for Efficient Electrical Stimulation of Human iPSC-Derived Cardiomyocytes. *Macromolecular Bioscience*, 2020, 19.6: e1900060 [MS-9096U]
<https://onlinelibrary.wiley.com/doi/abs/10.1002/mabi.201900060>
3. M. Kimura, et al, Increased mesodermal and mesendodermal populations by BMP4 treatment facilitates human iPSC line differentiation into a cardiac lineage. *Journal of Stem Cells & Regenerative Medicine*, 2019, 15.2: 45-51 [MS-90900]
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6971377/>

< Bone and Cartilage Research >

1. K. Endo, et al, Effect of Fibroblast Growth Factor-2 and Serum on Canine Mesenchymal Stem Cell Chondrogenesis. *Tissue Engineering Part A*, 2019, 25: 11-12 [MS-9096U]

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https://www.liebertpub.com/doi/10.1089/ten.TEA.2018.0177?url_ver=Z39.88-2003&rft_id=ori%3Arid%3Acrossref.org&rft_dat=cr_pub++0pubmed

2. T. Oshima, et al, A Scaffold-Free Allogeneic Construct From Adipose-Derived Stem Cells Regenerates an Osteochondral Defect in a Rabbit Model. *Arthroscopy: The Journal of Arthroscopic & Related Surgery*, 2019, 35.2: 583-593 [MS-9096U]
<https://www.sciencedirect.com/science/article/abs/pii/S0749806318307242>
3. K. Endo, et al, Comparison of the effect of growth factors on chondrogenesis of canine mesenchymal stem cells. *The Journal of Veterinary Medical Science*, 2019, 81.8: 1211-1218 [MS-9096U, M or V]
https://www.istage.jst.go.jp/article/jvms/advpub/0/advpub_18-0551/_pdf/-char/ja
4. A. Yamasaki, et al, Osteochondral regeneration using constructs of mesenchymal stem cells made by bio three-dimensional printing in mini-pigs. *Journal of Orthopaedic Research*, 2019, 37.6: 1398-1408 [MS-9096U]
<https://onlinelibrary.wiley.com/doi/abs/10.1002/jor.24206>
5. E. K. Breathwaite, et al, Scaffold-free bioprinted osteogenic and chondrogenic systems to model osteochondral physiology. *Biomedical Materials*, 2019, 14.6: 65010 [MS-9096U]
<https://iopscience.iop.org/article/10.1088/1748-605X/ab4243/meta>
6. S. Noda, et al, Effect of cell culture density on dental pulp-derived mesenchymal stem cells with reference to osteogenic differentiation. *Scientific Reports*, 2019, 9.1:5430 [MS-9096U, M or V]
<https://www.nature.com/articles/s41598-019-41741-w.pdf>
7. S. P. Grogan, et al, Cartilage tissue engineering combining microspheroid building blocks and microneedle arrays. *Connective Tissue Research*, 2019, 61(2) [MS-9096U]
<https://www.tandfonline.com/doi/full/10.1080/03008207.2019.1617280?scroll=top&needAccess=true>

< Vascular Research >

1. I. Pitaktong, et al, Early Vascular Cells Improve Microvascularization Within 3D Cardiac Spheroids. *Tissue Engineering Part C: Methods*, 2020, 26.2: 80-90 [MS-9096U]
<https://www.liebertpub.com/doi/abs/10.1089/ten.tec.2019.0228>
2. M. Itoh, et al, Development of an immunodeficient pig model allowing long-term accommodation of artificial human vascular tubes. *Nature Communications*, 2019, 10.1: 2244 [MS-9096U]
<https://www.nature.com/articles/s41467-019-10107-1>
3. K. Arai, et al, Cryopreservation method for spheroids and fabrication of scaffold-free tubular constructs. *PLoS ONE*, 2020, 15(4): :e0230428 [MS-9096U, V or M]
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0230428>
4. Y. Isshiki, et al, Co-Culture of a Brain Organoid Derived from Human iPSCs and Vasculature on a Chip. *2020 IEEE 33rd International Conference on Micro Electro Mechanical Systems (MEMS)*, 2019, 1024-1027 [MS-9096V]

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<https://ieeexplore.ieee.org/document/9056422>

< Infertility Research >

1. Y. Miyazaki, et al, Versican V1 in human endometrial epithelial cells promotes. *Reproduction*, 2019, 157.1: 53-64. [PrimeSurface dish/plate]
<https://rep.bioscientifica.com/view/journals/rep/157/1/REP-18-0333.xml>
2. T. Ishii, et al, Mild hypothermia promotes the viability of in vitro-produced bovine. *Journal of Reproduction and Development*, 2019, 65.3: 275-280. [MS-9096U]
https://www.istage.jst.go.jp/article/jrd/65/3/65_2018-142/article
3. E. S. Choi, et al, Effects of pyruvate and dimethyl- α -ketoglutarate, either alone. *Reproductive Medicine and Biology*, 2019, 18.4: 405-410. [MS-9096U]
<https://onlinelibrary.wiley.com/doi/10.1002/rmb2.12288>

< Immunotherapy >

1. N. Hiramatsu, et al, An analysis of monocytes and dendritic cells differentiated from human peripheral blood monocyte-derived induced pluripotent stem cells. *Medical Molecular Morphology*, 2020, 53: 63-72. [PrimeSurface dish/plate]
<https://link.springer.com/article/10.1007/s00795-019-00231-8>

< Transplantation and Cellular therapy >

1. H. Ueyama, et al, SLocal transplantation of adipose-derived stem cells has a significant therapeutic effect in a mouse model of rheumatoid arthritis. *Scientific Reports*, 2020, 10.1: 3076 [MS-9096V]
https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7033196/pdf/41598_2020_Article_60041.pdf

< Aggregation of iPSC >

1. Y. Kato, et al, Effect of liquid flow by pipetting during medium change on deformation of hiPSC aggregates. *Regenerative Therapy*, 2019, 12: 20-26. [MS-9096V]
<https://www.sciencedirect.com/science/article/pii/S2352320419300215>

< Others >

1. Y. Takahashi, et al. Therapeutic potential of spheroids of stem cells from human exfoliated deciduous teeth for chronic liver fibrosis and hemophilia a. *Pediatric surgery international*, 2019, 35.12: 1379-1388. [PrimeSurface dish/plate]
<https://link.springer.com/article/10.1007/s00383-019-04564-4>
2. M. Oka, et al. Exogenous cytokine-free differentiation of human pluripotent stem cells into classical brown adipocytes. *Cells*, 2019, 8.4: 373. [MS-9096V]
<https://www.mdpi.com/2073-4409/8/4/373>
3. D. Taniguchi, et al. Human lung microvascular endothelial cells as potential alternatives to human umbilical vein endothelial cells in bio-3D-printed trachea-like structures. *Tissue and Cell*, 2020, 63:

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101321. [MS-9096U]

<https://www.sciencedirect.com/science/article/abs/pii/S0040816619303660>

4. Y. Nakanishi, et al. Histological evaluation of tendon formation using a scaffold-free three-dimensional-bioprinted construct of human dermal fibroblasts under in vitro static tensile culture. *Regenerative therapy*, 2019, 11: 47-55. [MS-9096U, M or V]
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5. Y. Takeoka, et al. Regeneration of esophagus using a scaffold-free biomimetic structure created with bio-three-dimensional printing. *PLoS ONE*, 2019, 14.3: e0211339. [MS-9096U, M or V]
<https://journals.plos.org/plosone/article/file?type=printable&id=10.1371/journal.pone.0211339>
6. T. Kageyama, et al. Preparation of hair beads and hair follicle germs for regenerative medicine. *Biomaterials*, 2019, 212: 55-63. [MS-90350 and MS-9096U]
<https://reader.elsevier.com/reader/sd/pii/S0142961219302674?token=A5C61985425BDA2BB989D2155D089EEDA9F942639E3FAC653E381D1C02DF89ED8CED582D71F045A77E9DC2A05DF4C2A6>
7. K. Suzuki, et al. Directed differentiation of human induced pluripotent stem cells into mature stratified bladder urothelium. *Scientific reports*, 2019, 9.1: 1-13. [MS-9096V]
<https://www.nature.com/articles/s41598-019-46848-8>

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